مطالعه مقایسه‌ای ریخت‌شناسی، گرده‌شناسی و روغن‌های ضروری سه گونه از سرده شنبلیله (تیره باقلائیان) در ایران

رویا کرمیان* و زهرا حاج مرادی

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چکیده. سرده شنبلیله یکی از سرده‌های مهم تیره حبوبات است که بسیاری از گونه‌های آن ارزش تغذیه‌ای و دارویی دارند. در این پژوهش ویژگی‌های ریخت شناسی سه گونه از سرده شنبلیله (*T. disperma*، *T. subenervis* و *T. aphanoneura*) مورد بررسی قرار گرفت. روغن‌های ضروری حاصل از بخش-های هوایی دو گونه *T. aphanoneura* و *T. subenervis* با استفاده از GC و GC/MS مورد بررسی قرار گرفت و با گونه *T. disperma* مقایسه شد. اسپاتولنول (۱۵/۱٪) فراوان‌ترین جزء موجود در روغن ضروری گونه *T. subenervis* بود. همچنین روغن این گونه دارای مقدار اندکی از ژرماکرن D (۶٪) بود. روغن گونه *T. aphanoneura* نیز غنی از اسپاتولنول (۱۰/۴٪) بود. چهار ترکیب دیگر نیز کمتر از ۱٪ روغن این گونه را به خود اختصاص دادند، لیکن n- اوکتیل استات کمترین مقدار را نشان داد. بر طبق نتایج این تحقیق، داده‌های مورفولوژیکی و فیتوشیمیایی بهتر از داده‌های گرده، روابط گونه‌های مورد مطالعه را روشن می‌کند.

واژه‌های کلیدی. بقولات، روغن‌های ضروری، ریخت‌شناسی، گرده

Comparative study on the morphology, palynology and essential oil composition of three *Trigonella* L. species (Fabaceae) from Iran

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Abstract. *Trigonella* L. is one of the important genera of the family Fabaceae, many species of which have nutritional and medicinal properties. In this investigation, morphological characters related to habit and pollen grain of three Trigonella species i.e., *T. disperma*, *T. subenervis* and *T. aphanoneura* were studied. Oils from the aerial parts of two species were investigated by GC and GC/MS methods and compared with *T. disperma*, which had been studied in advance. *T. subenervis* represented oils rich in spathulenol (15.1%). It also contained a small amount of germacrene D (0.6%). *T. aphanoneura* also represented oils rich in spathulenol (10.4%). The amount of four compounds in this species was ≤1%, and n-octyl acetate was the lowest component in quantity. According what we found out, phytochemical and morphological data could clarify the relationships among the *Trigonella* species better than pollen morphological data.

Keywords. essential oil composition, Leguminosae, morphology, pollen, *Trigonella*

INTRODUCTION

Trigonella L. is a large genus with approximately 135 species, belonging to the family Fabaceae (Kawashty *et al.*, 1998). In Iranian plateau, the genus is represented by 58 annual and perennial species grouped into 12 sections (Boissier, 1872; Janighorban, 2004; Townsend, 1974). Many of the perennial species are endemic to Iran and belong to sect. *Ellipticae* (Rechinger, 1984). Various species of the genus are important for their medical and culinary properties (Abdel-Barry *et al.*, 2000; Chopra *et al.*, 1956; Giradon *et al*., 1985; Rapior & Bessiere, 2000; Sood, 1975).

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Table 1. The collections of three *Trigonella species.*

Species	Voucher specimen	Locality	Date	Altitude (m)
T. disperma Vassilcz.	BASU 11089	Kordestan: Ghorveh, after Hamekasi	26.06.2010	1700
T. aphanoneura Rech.f.	BASU 7222	Bakhtiary: Boroujen protected area	30.02.2010	1100
T. subenervis Rech.f.	BASU 14337	Khorassan: 15 km after Rivash, 80 km to Neyshabour	04.07.2010	1700

One of them, *T. foenumgraecum* L. is commonly grown as a vegetable (Grossheim, 1945; Hamzehee, 2000; Hedge, 1970). Many bioactive compounds such as protodioscin, trigoneoside, diosgenin, yamogenin have been isolated from fenugreek seeds (Giradon *et al*., 1986; Kabilan *et al*., 2002; Murakami *et al*., 2000; Pandita *et al*., 1991; Small *et al*., 1990; Yoshikawa *et al*., 1992). The important chemical constituents are saponins, coumarin, fenugreekine, nicotinic acid, phytic acid, scopoletin and trigonelline (Ullah Khan *et al*., 2009).

Studies on the pollen grains of leguminous plants (Brookes & Small, 1988; Clarke & Kupicha, 1976; Diez and Ferguson, 1994; Ferguson, 1990; Ferguson & Skvarla, 1981; Ferguson & St-riton, 1993; Hughes, 1997; Kovach 1985-2002; Small & Bassett, 1981) dealt mainly with the description of the pollen grains of certain genera, e.g. *Trigonella*, or tribes. However, there are only two reports on the oil composition in the genus dealt with *T. foenum-graecum* and *T. disperma* (Ahmadiani *et al*., 2004; Ranjbar *et al*., 2009).

In this investigation three *Trigonella* species, *i.e. T. disperma, T. subenervis* and *T. aphanoneura*, were studied from different aspects of morphology, pollen morphology and essential oil composition, and the relationships between the species were discussed. These species are endemic to Iran and belong to sect. *Ellipticae*. We report here the composition of the essential oils of *T. aphanoneura* and *T. subenervis* for the first time and compare them with those of *T. disperma.* The aim of the present study was to investigate whether or not the secondary compounds could be used as discriminative taxonomic markers in this group, especially between the closely related taxa.

MATERIAL AND METHODS

Plant material

Fresh materials of each species were sampled from nature. The locations of the collected species are given in Table 1. The voucher specimens were deposited at the Bu-Ali Sina University Herbarium (BASU), Hamedan, Iran.

Morphology

In morphological study, 28 qualitative and quantitative characters (Table 2) related to reproductive and vegetative organs were studied in 10 individuals of each species. Data were entered in Microsoft Excel. The spread sheet was later transformed into a file format suitable for phenetic analysis. Then, the principal coordinate analysis (PCO) was carried out using MVSP version 3.1, with a matrix of standardized data (Adams, 2001).

Essential oil composition

The air-dried aerial parts of the plants were subjected to hydrodistillation using a Clevenger-type apparatus for 3 h. Prior to the analysis, the oils obtained were dried over anhydrous sodium sulfate and stored in sealed glass vials at 4c. The oil was analyzed by GC and GC/MS.

GC analysis was performed on a Thennoquest-Finnigan Trace GC instrument equipped with FID detector and a capillary fused silica column (60 m×0.25 mm ID, coated with DB-l, film thickness, 0.25 μm).

The column temperature was kept at 60°C for 3 min and then programmed to 250°C at a rate of 5°C/min and then kept constant at 250°C for 10 min. Injector and detector temperatures were 250°C and 280°C, respectively. The flow rate of N_2 (carrier gas) was 1.1 ml/min; split ratio, 1:50. GC/MS analysis was carried out on a Thennoquest-Finnigan Trace GC-MS system equipped with a fused silica capillary column;
(60 m × 0.25 mm ID, coated with DB-l, film thickness, 0.25 μm) and interfaced with a quadruple detector.

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Table 2. Mean value of morphological features and character state matrix of three *Trigonella* species.

Morphological features	*T. subenervis*	*T. aphanoneura*	*T. disperma*
Stem length (cm)	26±0.85	23.5±0.88	45±1.05
Internode length (cm)	0.36±3	2.2±0.61	2.7±0.50
Leaf length (mm)	10.5±0.15	6±0.35	10.5±0.22
Leaflet length (mm)	7.5±0.04	6±0.11	7±0.08
Leaflet width (mm)	7±0. 07	4.5±0.01	5±0.02
Petiole length (mm)	8±0.04	4±0.03	8.5±0.12
Stipule length (mm)	3±1.88	3±2.31	4±2.60
Stipule width (mm)	0.5±0.32	1±0.26	0.7±0.18
Peduncle length (cm)	2.3±0.05	3.4±0.21	2.6±0.09
Calyx tube length (mm)	3±0.05	2.5±0.06	2.5±0.10
Calyx tooth length (mm)	2±0.05	1±0.05	1±0.10
Calyx length (mm)	5±1.10	3.5±1.10	3.5±0.90
Standard length (mm)	10±0.08	7±0.09	10±0.09
Standard width (mm)	7±1.23	7±1.51	8±1.08
Standard claw length (mm)	2±1.38	1±1.60	2±1.29
Wing length (mm)	10±0.09	7±0.06	9±0.04
Wing width (mm)	2±2.05	2±2.14	3±1.98
Keel length (mm)	9±0.70	7.5±0.72	8±0.55
Number of flowers	4±1.13	8±1.65	7±1.41
Pedicel length (mm)	3±0.11	3.7±0.11	3.5±0.10
Leaflet beak length (mm)	0.6±0.46	0.6±0.21	0.4±0.08
Pod length (mm)	10±0.76	16.5±0.88	12.5±0.68
Pod width (mm)	2.5±1.06	3.5±1.11	5±1.24
Pod stipa length (mm)	0.5±0.06	2±0.08	1.5±0.12
Seed number	1±1.01	3±0.98	2±0.86
Hair density on bract (dense = 1, glabrous = 0)	0	1	0
Fruit wing (absent = 0, present = 1)	0	0	1
Standard color (yellow = 0, violet = 1)	1	0	0
Standard form (spheroid = 0, obovate = 1)	0	0	1

The column temperature was kept at 60°C for 3 min and then programmed to 250°C at a rate of 5°C/min and then kept constant at 250°C for 10 min. Injector and transfer line temperatures were 250°C. The flow rate of Helium (carrier gas) was 1.1 ml/min; split ratio, 1:50.

The MS operating parameters were as follows: ionization energy, 70 eV; scan time, 1 s; mass range, 45-465 amu; ionization current, 150 µA.

The components of the oil were identified by their retention times, retention indices (RI) relative to C_6-C_{24} *n*-alkanes (on DB-1 column under the same conditions) and their mass spectra with those reported in literature and by computer matching with the WILEY 7.0 library as well as whenever possible, co-injection with authentic compounds available (Jennings & Shi-bamoto, 1980).

The percentage composition of the identified compounds was computed from the GC peak area without any correction factor and calculated relatively. The data related to oil compositions were analyzed with MVSP ver. 3.1 to understand the relationships between the species (Adams, 2001).

Pollen morphology

For light microscopy (LM), pollen samples were obtained from herbarium specimens and prepared using the standard method described by Erdtman (1960). Pollen grains were mounted on unstained glycerin jelly and observed with a Nikon type-2 microscope. Measurements were conducted on the basis of 25 readings from each 10 specimens. Polar axis (P), equatorial diameter (E), colpus length (L), colpi interval in granule site (G) and in non-granule site (NG), and shape index (P/E) were measured.

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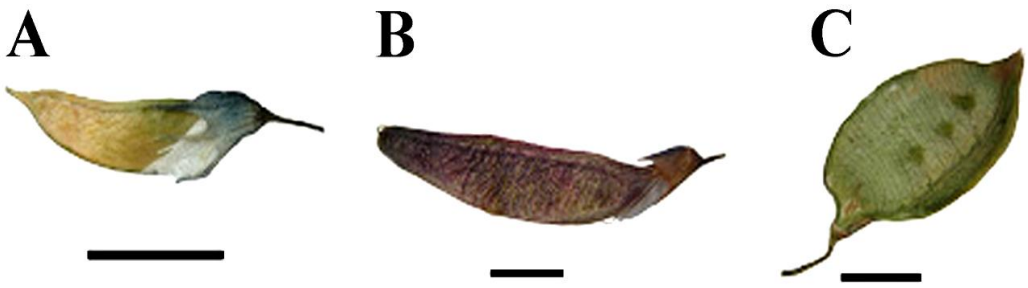


Fig. 1. Pods. **A**: *T. subenervis*; **B**: *T. aphanoneura*; **C**: *T. disperma* (with wing). Scale bar = 5 mm.

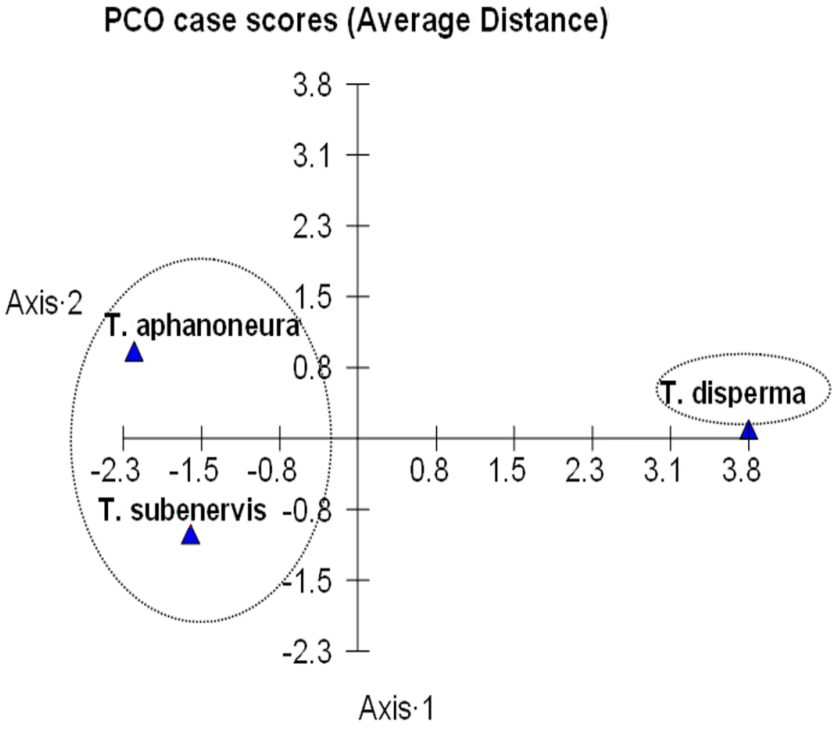


Fig. 2. PCO analysis of three *Trigonella* species based on morphological characters.

PCO analysis was performed using average distances in order to show phenetic relationships between the species based on pollen morphological characters.

RESULTS AND DISCUSSION

Morphology

This study reports the taxonomic relationnships between three *Trigonella* species armed with the knowledge about morphology, pollen morphology and oil composition. Three species studied were similar in morphology, but some characters related to pods separated them. Pod characters can be useful when the plants are in reproductive stage, but distinguishing among three species in their vegetative stage is so difficult. *T. disperma* is differentiated from other species by its longer pods (± 5 mm) with wings 1-1.5 mm in width (Fig. 1). The results from PCO analysis of the species based on morphological characters are illustrated in Figure 2.

Chemical composition of essential oils

The chemical composition of the essential oils of *T. subenervis* and *T. aphanoneura* were analyzed by means of GC/MS. The GS-separated compounds were identified by the recorded mass spectra in comparison with the mass spectra from the GS library. Table 3 gives the oil composition of representative samples of the three *Trigonella* species. In total, 23 compounds were identified in *T. subenervis*, *T. aphanoneura* and *T. disperma*, amounting to 41.3%, 40.6% and 87% of the oils, respectively.

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Table 3. Compounds identified in the aerial parts oil of three *Trigonella* species.

Compound[a]	Ri[b]	*T. subenervis* (%)	*T. aphanoneura* (%)	*T. disperma* (%)
p-Xylene	800	-	-	1.0
Decanal	1184	-	-	0.6
n-Octyl acetate	1190	1.3	0.4	-
Thymol	1265	1.5	0.9	1.1
Neryl acetate	1431	0.9	-	0.7
β-Lonene	1468	-	-	2.9
Germacrene D	1483	0.6	0.8	-
Pentacosane	1498	-	-	27.3
Nerolidol	1574	0.8	-	-
Spathulenol	1572	15.1	10.7	17.8
Caryophyllen oxide	1580	-	-	7.9
Hexadecane	1597	-	-	3.9
NI[c]	1661	6.7	11.2	5.3
NI[c]	1667	1.5	1.9	1.4
Heptadecane	1699	-	-	2.4
NI[c]	1708	3.2	4.8	3.1
NI[c]	1716	2.1	5.1	1.8
NI[c]	1728	1.2	6.0	1.3
Myristic acid	1738	1.8	3.1	2.1
Octadecane	1799	-	-	1.2
Hexahydroxyfarnesyl acetone	1829	5.8	-	6.7
Nonadecane	1898	-	0.6	0.9
Dibutyl phthalate	1920	2.7	1.8	3.9
Palmitic acid	1938	0.9	-	1.8
Farnesyl acetate	1990	1.3	1.0	-
Phytol	2100	-	-	4.0
Tricosane	2297	1.1	-	0.9
Tetracosane	2389	1.8	1.2	-
Total		50.3	49.5	100

[a]Compounds listed in order of their RI.

[b]RI (retention index) measured relative to *n*-alkanes (C_6-C_{24}) on DB-1 column.

[c]NI, not identified.

The oil composition of *T. aphanoneura* was found to be more similar to *T. subenervis* than *T. disperma*. Most of the compounds were volatile esters. Ranjbar *et al*. (2009) reported that in *T. disperma*, 87.1% of the oil comprised a total of 18 compounds. The major constituents of the oil were pentacosane (27.3%) and spathulenol (17.8 %). Decanal (0.6%) and neryl acetate (0.7%) were observed to have the lowest percentages in the oil. It was found that spathulenol had a high amount in both *T. subenervis* and *T. aphanoneura* (Table 3).

Some of the compounds such as one compound with high amount in *T. aphanoneura* (11.2%), were not identified. In addition, *n*-octyl acetate (0.4%) and germacrene D (0.6%) had the lowest amounts of *T. subenervis* and *T. aphanoneura*., respectively.

T. disperma showed major differences in its oil composition in comparison to other species. Its oil includes some compounds such as *p*-xylene, decanal, caryophyllen oxide, hexadecane, heptadecane, octadecane and phytol, which are all absent in other studied species.

In contrast, germacrene D and farnesyl acetate were not observed in *T. disperma*. A detailed list of the oil components of the species is given in Table 3 and PCO analysis based on oil composite-on data is illustrated in Figure 3. These phytochemical data verified morphological results and the separation of the species well. To sum up, these species synthesized many similar compounds in their essential oils.This fact can be justified by their similar morphology (biochemical convergence).

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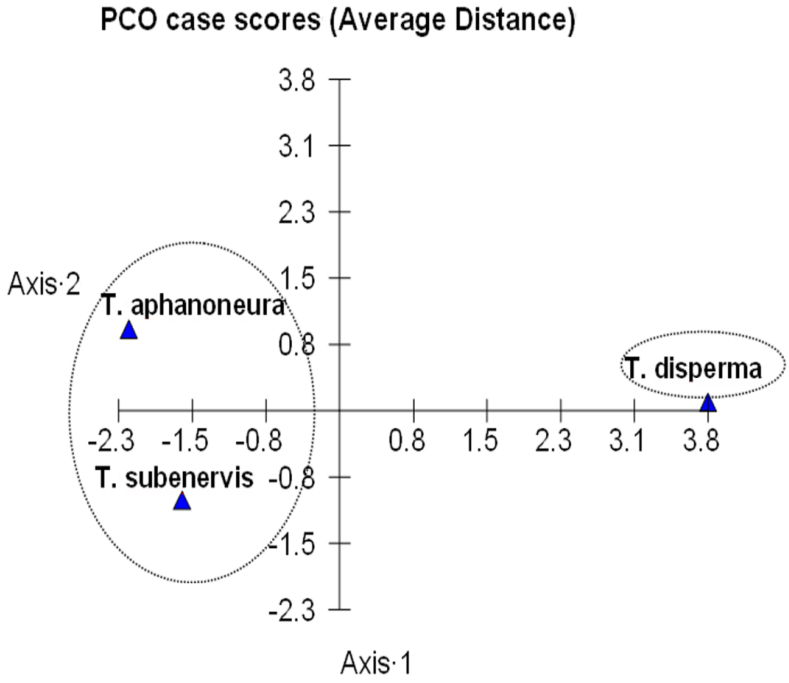


Fig. 3. PCO analysis of three *Trigonella* species based on leaf oil composition data.

Table 4. Taxa examined with measurements (µm) of the mean and ranges for pollen features of three *Trigonella*.

Species	E	P	G	NG	L	P/E
T. disperma	22(24±0.28)28.5	29(30.1±0.21)31	17(17.3±0.0.08)20	13(14.3±0.0.20)17	24(25.1±0.76)26	1.34
T. aphanoneura	21.3(22±0.46)24	28(28.5±0.15)31	17(18±0.11)20	13(13.9±0.18)15	24(25.0±0.89)26	1.33
T. subenervis	20(21.5±0.53)23	27(27.9±0.19)29	15(16.4±0.13)17	12(12.1±0.26)13	22(22.9±0.91)24	1.29

E: Equatorial diameter; **P**: Polar axis; **L**: Colpus length; **G** and **NG**: Colpi interval in granule site and non-granule site, respectively; **P/E**: Shape index. Means in parenthesis were expressed as ± SE.

However, the oil of *T. disperma* differs from the other two species significantly. PCO analysis verified the results of morphological study and showed that the differences referred to some constituents. In addition, the taxonomic distance of the species could be confirmed by chemical data.

Pollen morphology

The morphology of pollen grains represents more evidence which can be used to clarify the relationships between the species studied. The pollen grains of the tribe Trifolieae are radially symmetrical, isopolar trizonocolporate, with perforate tectume and varying shape and sculpture. The pollen morphology of this tribe shows that Trifolieae is a eurypalynous tribe (Gamal, 2006; Small & Bassett, 1981). In sect. *Ellipticae*, pollen grains are elliptic to oblong with often perforate ornamentation. Some features are common to several species of the section, while others are unique (Ranjbar *et al.*, 2012). The measurements of six pollen characters of the taxa studied are shown in Table 4, and the light micrographs are presented in Figure 4. The results of PCO analysis, based on pollen morphology, are not in agreement with morphology and phytochemistry (Fig. 5). In contrast to the phytochemistry and morphology-based analyses, *T. subenervis* is separated from other species because of its smaller pollen grains (Table 4).

CONCLUSION

According to what we found out, phytochemical and morphological data from our study could clarify the relationships among the *Trigonella* species better than pollen morphological data.

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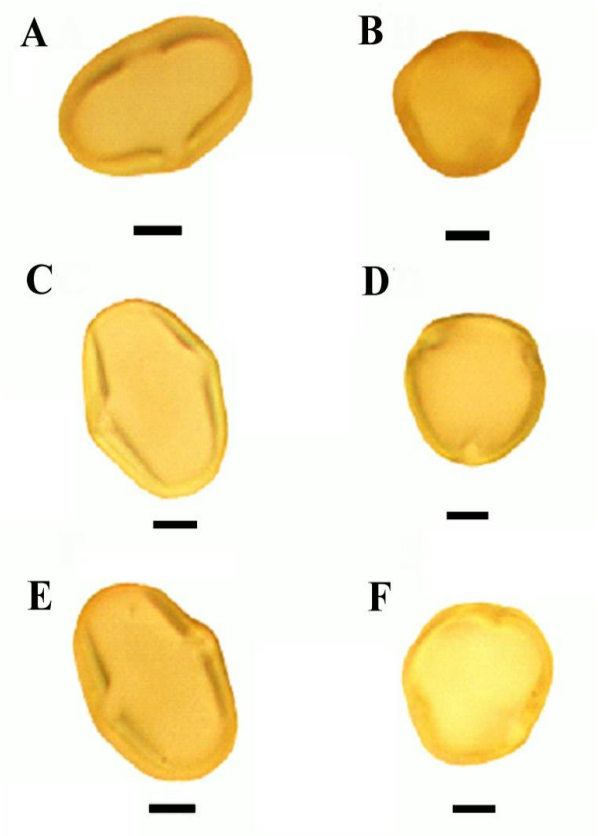


Fig. 4. LM polar and equatorial views of the pollen grains in three *Trigonella* species: **A, B**: *T. aphanoneura*; **C, D**: *T. disperma*; **E, F**: *T. subenervis*. Scale bars = 6 µm.

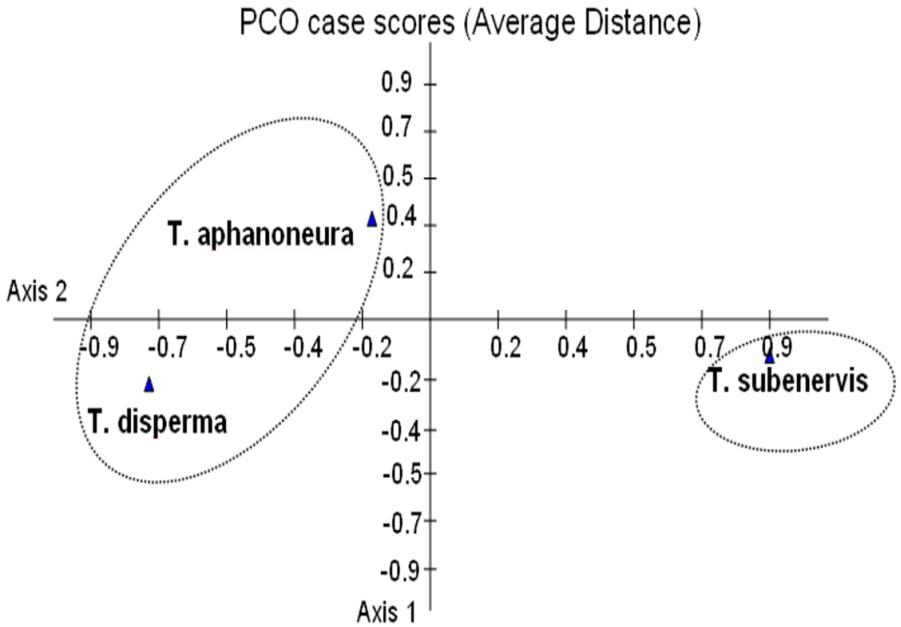


Fig. 5. PCO analysis of three *Trigonella* species based on pollen morphological characters.

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[DOI: 10.21859/acadpub.nbr.3.2.136]

REFERENCES

Abdel-Barry, A., Abdel-Hassan, J.A., Jawad, A.M. and Al-Hakim, A.H. 2000. Hypoglycemic effect of aqueous extract of the leaves of *Trigonella foenum-graecum* in healthy volunteers. – East Mediterr. Health J. 1: 83-88.

Adams, R.P. 2001. Identification of essential oil components by Gas Chromatography/Quadruple Mass Spectroscopy. – Allured Publishing Corporation, Illinois, USA, 3th Edition: 40-90.

Ahmadiani, A., Rustaiyan, A., Karamian, M., Kamalnejad, M.J. 2004. Volatile constituents from the oil of *Trigonella foenum-graecum* L. – J. Essent. Oil Res. 16: 1-2.

Al-Habori, M., Al-Aghbari, A.M. and Al-Mamary, M. 1998. Effects of fenugreek seeds and its extracts on plasma lipid profile: A study on rabbits. – Phytother. Res. 12: 572-575.

Boissier, E. 1872. *Trigonella* L. In: Flora Orientalis. 2: 65-91. – A. Asher & Co. B.V., Geneva. Amsterdam.

Brookes, B. and Small, E. 1988. Enhanced floral analysis by low temperature scanning electron microscopy. – Scanning Microsc. 2: 247-256.

Chopra, R.N., Nayyarand, S.L. and Chopra, I.E. 1956. Glossary of Indian Medicinal Plants. 1st Ed. – CSIR, New Delhi. 248 pp.

Clarke, G.C.S. and Kupicha, F.L.S. 1976. The relationships of the genus *Cicer* L. (Leguminosae): the evidence from pollen morphology. – Bot. J. Linn. Soc. 72: 35-44.

Diez, M.I. and Ferguson, L.K. 1994. The pollen morphology of the tribes Loteae and Coronilleae, *Lotus* L. and related genera (Papilionoideae: Leguminosae). – Rev. Palaeobot. Palynol. 81: 233-255.

Erdtman, G. 1960. The acetolysis method. – Sv. Bot. Tidskr. 54: 561-564.

Ferguson, I.K. 1990. The significance of some pollen morphology characters of the tribe Amorpheae and the genus *Mucuna* (tribe Phaseoleae). – Rev. Palaeobot. Palynol. 64: 129-136.

Ferguson, I.K. and Skvarla, J.I. 1981. The pollen morphology of the subfamily Papilionoideae (Leguminosae). – In: Polhill, R.M. and Raven, P. B. (eds.), Advances in legume systematics – Royal Bot. Gard. Kew: 859-896.

Ferguson, I.K. and Striton, C.H. 1993. Pollen morphology of the genera Leguminosae (Papilionoideae: Sophoreae), *Panurea* and *Bowdichia*. – Grana 2: 44-48.

Gamal, M.A.L. 2006. Comparative morphology of pollen grains of some taxa of tribe Trifolieae (Fabaceae: Papilionoideae) from Egypt. – Int. J. Bot. 2: 270-277.

Giradon, P., Bessiere, J.M., Baccou, J.C. and Sauvaire, Y. 1985. Volatile constitute of fenugreek seed. – Planta Medica 6: 533-534.

Giradon, P., Sauvaire, Y., Baccou, U.C. and Bessiere, J.M. 1986. Identification de Ia 3-hydroxy-4, 5-dimethyl-2 (SH)-furanone dans L'arome des graines de Fenugreek (*Trigonella foenum-graecum* L.). – Lebensmittel-Wissenschaft & Technologie 19: 44-46.

Grossheim, A.A. 1945. *Trigonella* L. In: Shishkin, B.K. (ed.), Flora USSR. 11: 102-129. – Akademiya Nauk SSSR, Moskva & Leningrad.

Hamzeh'ee, B. 2000. Some new and noteworthy plant records from Iran. – Iran. J. Bot. 8: 271-277.

Hedge, I.C. 1970. *Trigonella* L. In: Davis, P.H. (ed.), Flora of Turkey and the East Aegean Islands. 3:452-483. – Edinburgh University Press, Edinburgh.

Hughes, C.E. 1997. Variation in anther and pollen morphology in *Leucaena* Benth. (Leguminosae: Mimosoideae). – Bot. J. Linn. Soc. 123: 177-196.

Janighorban, M. 2004. A new record of the genus *Trigonella* (Fabaceae) for the flora of Iran. – Iran. J. Bot. 10: 177-179.

Jennings, W. and Shibamoto, T. 1980. Quantitative analysis of flavour and fragrance volatiles by Glass Capillary Gas Chromatography. – Academic Press, New York.

Kabilan, S., Kayalvizhi, A.R., Balasubramanian, S. and Ramalingan, C. 2002. Mass spectral analysis of solvent extracts of fenugreek seeds. – Asian J. Chem.14: 844-848.

Kawashty, S.A., Abdall, M.A., Gamal, E.L., Din, E.M. and Saleh, N.A.M. 1998. The chemosystematics of Egyptian *Trigonella* species. – Biochem. Syst. Ecol. 26: 851-859.

Kovach, W. 1985-2002. Institute of Earth Studies. University College of Wales, Aberystwyth, (Shareware), MVSP Version 3.2. – Kovach Computing Service.

Murakami, T., Kishi, A., Matsuda, H. and Yoshikawa, M. 2000. Medicinal foodstuffs. XVII. Fenugreek seed (3): structures of new furostanoltype steroid saponins, trigoneosides Xa, Xb, XIb, XIIa, XIIb, and XIIIa, from the seeds of Egyptian *Trigonella foenum-graecum* L. – Chem. Pharmaceutical Bull. 48: 994-1000.

Pandita, V.K., Nagarjan, S. and Sharma, D. 1991. Reducing hard seediness in fenugreek by scarification technique. – Seed Sci. Technol. 27: 627-637.

Ranjbar, M., Hajmoradi, Z. and Karamian, R. 2010. The taxonomic importance of leaf epidermis morphology and anatomy in *Trigonella disperma*. – TBJ. 1: 15-26.

Ranjbar, M., Karamian, R. and Hajmoradi, Z. 2009. Composition of the essential oil of *Trigonella disperma* from Iran. – Chem. Nat. Compod. 45: 116-117.

Ranjbar, M., Karamian, R., Hajmoradi, Z. and Joharchi, M.R. 2012. A revision of *Trigonella* sect. *Ellipticae* (Fabaceae) in Iran. – Nord. J. Bot. 30: 17-35.

Rapior, S. and Bessiere, J.M. 2000. The fenugreek odor of *Lactarius helvus*. – Mycologia 92: 305-

308.

Rechinger, K.H. 1984. *Trigonella*. In: Rechinger, K.H. (ed.): Flora Iranica. 157: 207-253. – Akad. Druck-und Verlagsanstalt, Graz.

Small, E., Bassett, J.I. and Crompton, W. 1981. Pollen variation in tribe Trigonelleae (Leguminosae) with special reference to *Medicago*. – Pollen et Spores 12: 295-319.

Small, E., Brookes, B. and Lassen, P. 1990. Circumscription of the genus *Medicago* (Leguminosae) by seed characters. – Canadian J. Bot. 68: 613-629.

Sood, A. 1975. Chemical compounds from the leaves of *Trigonella foenum-graecum* L. – Indian J. Pharmacol. 37: 100-101.

Townsend, C.C. 1974. – In: Townsend, C.C. and Guest, E. (eds.) Flora of Iraq 3: 86-112. – Ministry of Agriculture & Agrian Reform Republic of Iraq, Baghdad.

Ullah Khan, F., Durrani, F.R., Sutan, A., Ullah Khan, R. and Naz, S. 2009. Effect of fenugreek (*Trigonella foenum-graecum*) seed extract on visceral organs of broiler chicks. – J. Agriculture Biol. Sci. 4: 58-60.

Yoshikawa, M., Murakami, T., Komatsu, H., Murakami, N., Yamahara, J. and Matsuda, H. 1997. Medicinal food stuffs. IV. Fenugreek seed (1): structures of trigoneosides Ia, Ib, IIa, IIb, IIIa, and IIIb, new furostanol saponins from the seeds of Indian *Trigonella foenum graecum* L. – Chem. Pharmaceutical Bull. 4: 81-87.

*****

Karamian, R. and Hajmoradi, Z. 2016. Comparative study on the morphology, palynology and essential oil composition of three *Trigonella* L. species (Fabaceae) from Iran. – Nova Biol. Reperta 3 (2): 136-144.

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